Efficient in vivo priming of specific cytotoxic T cell responses by neonatal dendritic cells

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Abstract

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Reference


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In early life, a high susceptibility to infectious diseases as well as a poor capacity to respond to vaccines are generally observed as compared with observations in adults. The mechanisms underlying immune immaturity have not been fully elucidated and could be due to the immaturity of the T/B cell responses and/or to a defect in the nature and quality of Ag presentation by the APC. This prompted us to phenotypically and functionally characterize early life murine dendritic cells (DC) purified from spleens of 7-day-old mice. We showed that neonatal CD11c+ DC express levels of costimulatory molecules and MHC molecules similar to those of adult DC and are able to fully mature after LPS activation. Furthermore, we demonstrated that neonatal DC can efficiently take up, process, and present Ag to T cells in vitro and induce specific CTL responses in vivo. Although a reduced number of these cells was observed in the spleen of neonatal mice as compared with adults, this study clearly shows that neonatal DC have full functional capacity and may well prime Ag-specific naïve T cells in vivo. The Journal of Immunology, 2002, 168: 2219–2224.

The neonatal period is marked by a high susceptibility to infections. This susceptibility is associated with a marked limitation of the capacity of most vaccines to induce protective immune responses, as observed in neonates and young infants as well as in newborns from various animal species including mice (1). The understanding of immune immaturity is an important issue given the global burden of infectious diseases in early life. However, the mechanisms responsible for the limitations of infant immune responses are not yet fully elucidated. Although studies of neonatal T lymphocyte function showed several intrinsic defects such as low proliferation rate in response to anti-CD3 stimuli (2), Th2-biased response (3, 4), or weak expression of CD40 ligand on activation (5), recent findings suggest that this could be essentially attributed to the nature and quality of Ag presentation by professional cells, as demonstrated previously for neonatal macrophages (6). However, it is now well established that dendritic cells (DC) are the main APC with a unique ability to sensitize naïve T cells and induce primary immune responses (7). Human cord blood DC were recently shown to be less efficient than adult DC in supporting proliferation of T cells in response to antigenic stimulation (8). Because DC drive the development of Th1 response through the production of IL-12, which appears limited both in murine and human neonates, it was also suggested that the lack of Th1 response observed in neonatal mice could be due to suboptimal Th1 cell activation by neonatal DC (1). Recently, adherent cells (including DC) from spleens of mice 4 wk of age were shown to be defective in presenting tetanus toxoid to Ag-specific T cell clones (9). Altogether, these observations suggested that neonate DC may not be fully functional and could present a limited efficiency to induce Th1 and CTL activation. This prompted us to phenotypically and functionally characterize early life murine DC, including their ability to specifically activate CD8+ T cells in vivo. In this study, DC were purified from spleens of 7-day-old mice, which best corresponds to the stage of immune maturation of human newborns in terms of vaccine Ab responses and maturation of T cell responses (reviewed in Ref. 1). The number and the phenotype of these cells were characterized, and the function of these neonate DC was analyzed by determining their ability to mature, take up, process, and present Ag to CD8+ T cells, as well as their capacity to induce in vivo specific CTL responses.

**Materials and Methods**

**Mice**

BALB/c mice, originally obtained from CER Janvier (Le Genest St. Isle, France), were bred and housed onsite. Periodic screening showed the colony to be free of commonly occurring infectious agents. Breeding cages were checked daily for new births, and the day of birth was recorded as the day the litter was found. Pups were weaned, and males and females were separated at 3 wk of age. Neonatal animals were defined as those 7 days old, and adult mice used in this study were 8–10 wk old.

**Peptide and detoxified recombinant adenylate cyclase**

The synthetic peptide p118–126 (RPQASGYVM) carrying the 118–126 sequence from the lymphocytic choriomeningitis virus (LCMV) nucleoprotein corresponding to a class I and class II H-2d-restricted epitope (10, 11) was purchased from NeoSystem (Strasbourg, France). This peptide was tested for the presence of endotoxin using a chromogenic test (*Limulus* amebocyte lysate test; BioWhittaker, Fontenay-sous-Bois, France), and no endotoxin was detected.

The detoxified adenylate cyclase from *Bordetella pertussis* (CyaA-E5) and the CyaA-E5 bearing the LCMV epitope (CyaA-E5-LCMV) were gifts from D. Ladant (Pasteur Institute, Paris, France). Their production and purification have been previously described (12).

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Culture medium and cell line
Complete medium (CM) consisted of RPMI 1640 containing l-alanyl-l-glutamine dippeptide supplemented with 10% FCS, 5 × 10⁻⁵ M 2-ME, and antibiotics (penicillin, 100 U/ml, and streptomycin, 100 µg/ml).

The MV11H7 T cell hybridoma specific for the H-2⁵-restricted 118–126 LCMV epitope was generated in the laboratory as previously described (13).

DC purification
DC were purified from spleens of 7-day-old or 8- to 10-wk-old mice by positive selection according to CD11c expression using Automacs (Miltenyi Biotec, Bergisch Gladbach, Germany), following the manufacturer’s procedure. The purity of the two fractions was checked by flow cytometry on a FACSCalibur or FACScan (BD Biosciences, San Jose, CA), using a PE-conjugated anti-CD11c (HL3 clone; BD PharMingen, San Diego, CA) for labeling. The purity of the fraction was usually >97%.

Flow cytometry analysis
Cells were washed in PBS supplemented with 1% BSA and 0.1% NaN₃, incubated with anti-CD32/CD16 (2.4G2 clone; BD PharMingen) to avoid nonspecific binding, and labeled for 30 min at 4°C with the following Abs purchased from PharMingen: PE- or APC-conjugated anti-CD11c mAb (HL3 clone); PerCP-conjugated anti-CD3 mAb (17A2); PE-conjugated anti-CD4 mAb (RM4-5); FITC-conjugated anti-CD8 (53-6.7); PE-conjugated anti-CD11c (HL3 clone); PerCP-conjugated anti-CD80 mAb (16-10A1); FITC-conjugated anti-CD86 mAb (GL1); FITC-conjugated anti-I-Ad mAb (AMS-32.1). As negative controls, cells were stained with corresponding isotype-matched mAbs. Acquisition was performed on a FACSCalibur or FACScan (BD Biosciences) using a FACSDiva software.

CD8° T cells were purified from spleens of adult BALB/c or C57BL/6 mice by positive selection according to CD8α expression using Automacs (Miltenyi Biotec), following the manufacturer’s procedure. Then, 1 × 10⁶ cells were cocultured with various numbers of CD11c⁺ cells purified from spleens of 7-day-old or 8-wk-old BALB/c mice. After 72 h, [³H]thymidine (50 µCi/ml; ICN) was added to the wells, and the cells were harvested 6 h later with an automated cell harvester (Skatron). Incorporated thymidine was detected by scintillation counting. In all experiments, each point was done in duplicate. Results are expressed as the mean cpm. MLC assay
CD8° T cells were purified from spleens of adult BALB/c or C57BL/6 mice by positive selection according to CD8α expression using Automacs (Miltenyi Biotec), following the manufacturer’s procedure. Then, 1 × 10⁷ cells were cocultured with various numbers of CD11c⁺ cells purified from spleens of 7-day-old or 8-wk-old BALB/c mice. After 72 h, [³H]thymidine (50 µCi/ml; ICN) was added to the wells, and the cells were harvested 6 h later with an automated cell harvester (Skatron). Incorporate thymidine was detected by scintillation counting. In all experiments, each point was done in duplicate. Results are expressed as the mean cpm. Mouse immunization
Purified CD11c⁺ dendritic cells from spleens of 7-day-old or 8- to 10-wk-old BALB/c mice were incubated for 1 h with or without 50 µM p111–126 in Stem Stem Span medium (StemCell, Meylan, France). After extensive washing, 2 × 10⁶ cells were i.v. injected into 8- to 10-wk-old syngeneic mice.

In vitro cytotoxicity assay
Splenocytes from immunized mice were isolated 7 days after DC injection and restimulated in vitro during 5 days with p111–126 (0.1 µg/ml) in the presence of syngeneic irradiated naive spleen cells. The cytotoxic activity was determined in a 5-h in vitro ⁵¹Cr release assay as previously described (12). Briefly, P815 tumor cells incubated with or without the p111–126 Peptide (50 µM) were used as target cells. Various E:T ratios were used, and all assays were performed in duplicate. ⁵¹Cr release in each well was counted using a MicroBeta Trilux liquid scintillation counter (Wallac, Turku, Finland). Percentage of specific lysis was calculated as 100 × [experimental release − spontaneous release]/(maximal release − spontaneous release). Maximum release was obtained by adding 10% Triton X-100 to target cells, and spontaneous release was determined with target cells incubated without effector cells. Results are expressed as Δ% of specific lysis (% lysis in the presence of peptide − % lysis in the absence of peptide).

Results
Number and phenotypical characterization of neonatal DC
It is well established that functional splenic DC in adult mice express high levels of the CD11c marker. Thus, to analyze the function of neonatal DC, CD11c⁺ splenocytes were isolated by positive selection on MACS from spleens of 7-day-old BALB/c mice using magnetic CD11c microbeads. The number of cells recovered was compared with the number of DC isolated from adult BALB/c mice (Table I); 12- to 50-fold less CD11c⁺ cells were found into the spleen of 7-day-old mice as compared with spleen from adult mice. Indeed, 0.1–0.4 × 10⁶ CD11c⁺ cells were recovered in their spleens, corresponding to 0.15–0.7% of splenocytes, whereas 3.3% of cells expressed the CD11c molecule in the spleen of adult mice. However, DC:T cell and DC:B cell ratios in the spleens of 7-day-old and adult mice were quite similar. Expression of co-stimulatory molecules and MHC class II was also assessed on neonatal CD11c⁺ cells and was compared with the expression of these markers on adult DC (Fig. 1). Both neonatal and adult DC expressed similar levels of CD40, CD80, CD86, and MHC class II.

Table 1. Number and percentage of CD11c⁺ cells in spleens from 7-day-old and 8-wk-old BALB/C mice

<table>
<thead>
<tr>
<th>Splenotype</th>
<th>Total No. of Splenocytes (×10⁶)</th>
<th>No. of CD11c⁺ Cells (×10⁶)</th>
<th>% of CD11c⁺ Cells</th>
<th>DC:T Cell Ratio</th>
<th>DC:B Cell Ratio</th>
</tr>
</thead>
<tbody>
<tr>
<td>7-day-old mice</td>
<td>55–75</td>
<td>0.1–0.4</td>
<td>0.15–0.7</td>
<td>0.1</td>
<td>0.028</td>
</tr>
<tr>
<td>8-wk-old mice</td>
<td>150</td>
<td>5</td>
<td>3.3</td>
<td>0.08</td>
<td>0.04</td>
</tr>
</tbody>
</table>

*Number and percentage of CD11c⁺ cells in spleen were determined by flow cytometry. Data represent results obtained from 10 mice. DC:T and DC:B cell ratios were determined after flow cytometry analysis; DC are characterized as CD11c⁺ cells, T cells are characterized as CD3⁺ and CD8⁺ or CD4⁺, and B cells are characterized as B220⁺ cells. Data represent results obtained from five mice.*
molecules, indicating that expression of the main molecules involved in DC-T cell interaction at the surface of neonatal CD11c<sup>+</sup> DC was not affected.

**Ability of neonatal DC to mature following LPS stimulation**

DC represent a heterogeneous population of cells. Immature DC are characterized by an efficient capacity to capture Ag but a low efficiency to stimulate naive T cells. After Ag capture and activation by microbial or inflammatory stimuli, DC mature and acquire the ability to stimulate naive T cells. DC maturation is characterized by up-regulation of expression of costimulatory and MHC molecules at the surface of DC (7). Thus, to determine the ability of neonatal DC to mature, purified CD11c<sup>+</sup> splenocytes from 7-day-old BALB/c mice were stimulated in vitro with LPS. The level of expression of CD40, CD80, CD86, and I-A<sup>d</sup> molecules was determined by FACS analysis and compared with the expression level of these markers before stimulation. Fig. 2 shows that LPS induced a marked increase in the surface expression of CD40, CD80, CD86, and I-A<sup>d</sup> with the same efficiency than adult CD11c<sup>+</sup> cells. Furthermore, a high increase of MHC class II expression was observed for both neonatal and adult DC after LPS stimulation although this increase cannot be compared between these two populations due to very high levels of expression. Altogether, these results clearly show the full maturation potential of neonatal DC.

**Ag uptake by neonatal DC**

Immature DC are very efficient in Ag capture using several pathways and DC maturation process is associated with a loss of endocytic and phagocytic capacities (7). To compare the Ag uptake capacity between neonatal and adult DC, the endocytic activity of these cells was assessed by determining the internalization of FITC-conjugated dextran by immature and mature purified DC from spleens of 7-day-old and 8-wk-old BALB/c mice. Freshly purified DC were incubated at 37°C with FITC-dextran; then its internalization was assessed by flow cytometry on gated CD11c<sup>+</sup> cells. As controls, purified DC were incubated at 4°C to block endocytosis (Fig. 3). Immature neonatal and adult DC show similar efficiency in the uptake of FITC-dextran, demonstrating the efficient endocytic capacity of neonatal DC. As expected, after maturation following LPS stimulation, both neonatal and adult DC lose their endocytic capacity as shown by the similar profile obtained after incubation at 4°C and 37°C for both DC populations. These results clearly show the capacity of neonate DC to take up Ag and confirm the ability of these cells to mature after LPS stimulation.

**Ag processing and presentation by neonatal DC**

The ability of neonatal DC to present Ag was assessed by their capacity to stimulate the MV1H7 CD8<sup>+</sup> T cell hybridoma, specific for the L<sup>d</sup>-restricted epitope 118–126 from the nucleoprotein of LCMV. Purified CD11c<sup>+</sup> splenocytes from 7-day-old BALB/c mice were loaded with various concentrations of the p<sub>118–126</sub> synthetic peptide. These cells were used as APC to stimulate the
Freshly purified CD11c+ secreted by the stimulated hybridoma was determined by the CTLL-2 proliferation assay. Data represent means ± SD of duplicates and are expressed in cpm of incorporated [3H]thymidine. Results are representative of three experiments.

In vivo induction of specific CTL responses by neonatal DC in adult mice

Finally, the ability of neonatal DC to activate naïve T cells and to induce in vivo-specific CTL responses was determined. Because it was previously shown that DC reach the spleen after i.v. injection (15), induction of CTL responses by neonatal DC was assessed in the spleen after i.v. immunization with peptide-pulsed DC. Freshly purified CD11c+ splenocytes from 7-day-old and 8-wk-old BALB/c mice were pulsed with P118–126. Then, 2–3 × 10^5 DC were injected i.v. to adult syngeneic mice. Seven days after the injection, splenocytes from immunized mice were stimulated in vitro with P118–126. Five days later, CTL activity was assessed by 51Cr release assay using H-2d P815 target cells pulsed or not with the same peptide. As shown in Fig. 6, neonatal DC loaded with P118–126 and injected into adult mice induced peptide-specific CTL responses comparable to the ones induced by adult DC, although a heterogeneity in CTL activity was observed between the different panels.
mice tested. The low CTL responses observed after injection of neonatal and adult mice unloaded DC demonstrated the specificity of the CTL responses induced by peptide-pulsed DC. These results clearly demonstrate that in vitro pulsed neonatal DC are able to induce in vivo-specific CTL responses and thus are able to prime naive CD8<sup>+</sup> T cells in vivo.

Discussion

Although newborns are at risk of exposure to many infectious diseases, the immaturity of the neonatal immune system leads to postdine most vaccinations to 2–3 mo of age. The mechanisms responsible for such limited neonatal immune responses are not fully elucidated. The neonatal murine T cell compartment seems functional because CTL and Th1 responses can be induced in neonates after immunization using strong Th-1 driving adjuvants or delivery systems (16–20). Some evidences suggest that the lack of appropriate immune T cell responses of neonates is due to the nature and quality of Ag presentation (1). The aim of this study was thus to determine the potential role of these cells in impaired neonatal immunity. For this, we analyzed the properties of DC purified from spleens of 7-day-old mice, the age which best corresponds to the main known characteristics of human neonatal immune maturation (1). Neonatal DC were pulsed according to the expression of the CD11c molecule which is expressed on both immature and mature DC in the spleen from adult mice. We found that this population is 12 to 50 fold less represented in neonatal spleen as compared with adult spleen. In contrast to what has been observed for neonatal macrophages (6), neonatal DC express similar levels of MHC class II and costimulatory molecules as adult DC. Moreover, our study clearly showed that neonatal CD11c<sup>+</sup> DC are functional and are able to induce in vivo CD8<sup>+</sup> T cell responses. Indeed, these cells were capable of full maturation after LPS activation. Evaluation of their Ag-presenting ability showed that these APC can process and present Ag to a CD8<sup>+</sup> T cell hybridoma and induce proliferation of allogeneic CD8<sup>+</sup> T cells in MLR assay as efficiently as adult DC. Finally, high specific cytotoxic activities were induced in vivo by in vitro pulsed neonatal DC injected to adult mice, showing the capacity of these cells to prime in vivo Ag-specific naive T cells. These last observations are in accordance with a recent paper showing that APC from 3- and 7-day-old mice induce a strong MLR from allogeneic CD8<sup>+</sup> T cells, comparable to the response obtained with adult stimulator cells (9) indicating the potentiality of neonatal DC to activate CD8<sup>+</sup> T cells. However in the same study, the authors showed a defect of neonatal APC to activate CD4<sup>+</sup> T cells due to a low expression of MHC class II molecules. By contrast, we demonstrate here that CD11c<sup>+</sup> neonatal DC express normal level of MHC class II molecules. This discrepancy could be mainly explained by difference of purification procedures used in both studies. Indeed, using cell density and adherence selective criteria, Muthukumar et al. (9) isolated 1.7 × 10<sup>5</sup> cells per spleen from 7-day-old mice among which only 7% were CD11c<sup>+</sup> cells (corresponding to 0.12 × 10<sup>5</sup> cells from one neonatal spleen), whereas in our study, using CD11c microbeads, 4 × 10<sup>5</sup> CD11c<sup>+</sup> cells were purified per neonatal spleen (i.e., 33-fold more than in the Muthukumar’s study). This strongly suggests that cell density and adherence criteria mainly select nonfunctional progenitor cells rather than functional DC present in neonatal spleen. Thus, it is of particular interest to reinvestigate the CD4<sup>+</sup> T cell response induced by in vitro-pulsed neonatal DC expressing the CD11c<sup>+</sup> molecule to determine whether this population is really defective in inducing Th response. Furthermore, because a preferential Th2 polarization of early life responses is observed in vivo (4, 21), the Th polarization induced by neonatal CD11c<sup>+</sup> DC has to be determined.

Altogether, our results indicate that the weak Th1/CTL immune responses observed in neonates cannot be explained by an intrinsic defect in nature and quality of Ag presentation by APC, at least for the CD8<sup>+</sup> T cell responses. This limitation is also unlikely to be due to the reduced number of DC found in neonatal spleens, because the DC:T cell ratio was 0.1 as compared with 0.08 in adult spleens, suggesting that their representation is sufficient for APC-T cell interactions to take place. It has been suggested that the lack of CTL induction in neonate could be due to the biased Th2 response observed in neonates (1), and the contribution of neonatal DC to this preferential polarization remains to be determined. We previously showed that both Th and CTL responses are induced in

![FIGURE 5.](http://www.jimmunol.org/)

**FIGURE 5.** Effective in vitro priming of allogeneic T cells by neonatal DC. A fixed number of purified H-2<sup>B</sup> CD8<sup>+</sup> T cells obtained from C56BL/6 mouse spleens was cocultured during 3 days with various numbers of H-2<sup>B</sup> CD11c<sup>+</sup> cells isolated from 7-day-old or 8-wk-old BALB/c mice. As controls, purified H-2<sup>B</sup> CD8<sup>+</sup> T cells obtained from BALB/c mice were cocultured with H-2<sup>B</sup> CD11c<sup>+</sup> cells from 7-day-old or 8-wk-old BALB/c mice. Then, a proliferation assay was performed. Data represent means ± SD of duplicates and are expressed in cpm of incorporated [3H]thymidine. Results are representative of three experiments.

![FIGURE 6.](http://www.jimmunol.org/)

**FIGURE 6.** Neonatal DC induce in vivo strong specific CTL response. CD11c<sup>+</sup> cells were isolated from 7-day-old or 8-wk-old BALB/c mice and were pulsed with P<sub>118-126</sub> and transferred i.v. into 8- to 10-wk-old syngeneic mice. As controls, some mice were injected with unpulsed DC from 7-day-old or 8- to 10-wk-old BALB/c mice. Seven days later, mice were sacrificed, and splenocytes were restimulated in vitro with P<sub>118-126</sub> and irradiated syngeneic splenocytes. CTL activities were tested on <sup>51</sup>Cr-labeled P815 target cells pulsed or unpulsed with P<sub>118-126</sub>. Results show the percentage of specific lysis obtained with P815 pulsed with the peptide corrected for background lysis obtained with unpulsed P815 target cells at an E:T ratio of 90. Background lysis was below 15% under all conditions. Each point corresponds to an individual mouse; mean values of cumulative data from separate experiments. Results obtained between neonatal and adult DC are not statistically different using a t test (p = 0.72).
adult mice immunized with DC loaded with \( p_{118-126} \), which contains both H-2\(^d \) class I and class II epitopes (22). The comparable LCMV-specific CTL responses induced by either neonatal or adult DC do suggest that the Th response induced by \( p_{118-126} \)-pulsed neonatal DC does not negatively affect the CTL response. Thus, this demonstration of the full potential capacity of neonatal murine DC to induce strong CTL responses in vivo makes them an attractive target for vaccination strategies to induce protective immunity in early life.

Acknowledgments

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References