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Reference

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A Novel Tumor Necrosis Factor–mediated Mechanism of Direct Epithelial Sodium Channel Activation

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Abstract

Rationale: Alveolar liquid clearance is regulated by Na+ uptake through the apically expressed epithelial sodium channel (ENaC) and basolaterally localized Na+–K+–ATPase in type II alveolar epithelial cells. Dysfunction of these Na+ transporters during pulmonary inflammation can contribute to pulmonary edema.

Objectives: In this study, we sought to determine the precise mechanism by which the TIP peptide, mimicking the lectin-like domain of tumor necrosis factor (TNF), stimulates Na+ uptake in a homologous cell system in the presence or absence of the bacterial toxin pneumolysin (PLY).

Methods: We used a combined biochemical, electrophysiological, and molecular biological in vitro approach and assessed the physiological relevance of the lectin-like domain of TNF in alveolar liquid clearance in vivo by generating triple-mutant TNF knock-in mice that express a mutant TNF with deficient Na+ uptake stimulatory activity.

Measurements and Main Results: TIP peptide directly activates ENaC, but not the Na+–K+–ATPase, upon binding to the carboxy-terminal domain of the α subunit of the channel. In the presence of PLY, a mediator of pneumococcal-induced pulmonary edema, this binding stabilizes the ENaC-PIP2-MARCKS complex, which is necessary for the open probability conformation of the channel and preserves ENaC-α protein expression, by means of blunting the protein kinase C-α pathway. Triple-mutant TNF knock-in mice are more prone than wild-type mice to develop edema with low-dose intratracheal PLY, correlating with reduced pulmonary ENaC-α subunit expression.

Conclusions: These results demonstrate a novel TNF-mediated mechanism of direct ENaC activation and indicate a physiological role for the lectin-like domain of TNF in the resolution of alveolar edema during inflammation.

Keywords: epithelial sodium channel; pneumonia; protein kinase C-α; pulmonary edema; tumor necrosis factor
Alveolar liquid clearance (ALC) is critical to preventing excess fluid accumulation in the alveoli (alveolar edema), which compromises gas exchange (1–4). Dysfunctional ALC correlates with morbidity and mortality in ARDS patients (5). ALC is mediated mainly by vectorial Na⁺ transport through the apical epithelial sodium channel (ENaC) and the basolateral Na⁺-K⁺-ATPase in type II alveolar epithelial cells (AECs). The magnitude of ENaC-mediated Na⁺ uptake in type II AEC depends on (i) proteolytic cleavage-induced excision of inhibitory tracts within proximal regions of the extracellular domains of the α and γ subunits, (2) the average open probability (Pₒ) time of the channel, and (3) the expression level of functional channels. Functional ENaC channels typically consist of α, β, and γ subunits (6–8), with the α subunit being crucial for neonatal and adult lung liquid clearance (9). ENaC subunit expression is regulated by clathrin-mediated endocytosis (10), as well as by Nedd-4-1- or -2-mediated ubiquitinylation and degradation (11). The open conformation of ENaC is stabilized by its direct binding to phosphatidylinositol 4,5-bisphosphate (PIP2) in complex with myristoylated alanine-rich C kinase substrate (MARCKS) (12–14). MARCKS has been demonstrated to be a target of both the α and δ isoforms of protein kinase C (PKC-α and PKC-δ, respectively) (15, 16).

Proinflammatory cytokines, including IL-1β, transforming growth factor β, and tumor necrosis factor (TNF) can significantly impair ENaC activity, in a p38 MAP kinase–or PKC-dependent manner (17–20). TNF has a dual role in ENaC regulation (21). Activation of TNF receptor 1 mediates transcriptional inhibition of all three ENaC subunits—α, β, and γ—and induces PKC-dependent post-translational inhibition of ENaC-α subunit expression in vivo and in damaged lungs in vivo (19). By contrast, TNF increases edema reabsorption in a rat pneumonia model (22) and stimulates Na⁺ uptake in A549 cells in a catecholamine-independent manner (23). These activities are most likely mediated by the lectin-like domain of the cytokine (24), which is spatially distinct from the receptor binding sites (25, 26). The lectin-like domain of TNF is mimicked by the 17-amino acid circular TIP peptide (26), which increases amiloride-sensitive Na⁺ uptake in type II AECs (27, 28). The TIP peptide has been shown to activate ALC in several animal models of hydrostatic and permeability edema (21, 29–31). In a recently performed, placebo-controlled, double-blind Phase I clinical trial, TIP peptide (also named AP301) was observed not to induce serious adverse effects upon inhalation (32).

Bacterial and viral toxins have been shown to induce significant changes in ENaC Pₒ and/or expression, leading to impaired ALC (33–36). It is therefore of high clinical relevance to identify novel mechanisms that increase ENaC function during pulmonary edema associated with ARDS and pneumonia, for which no standard therapy exists to date. Preliminary data from a Phase Ia interventional, randomized, double-blind, placebo-controlled, parallel-group study of patients with acute lung injury indicated that oral inhalation of TIP peptide caused earlier onset and more pronounced activation of pulmonary edema clearance, as compared to placebo-treated patients (Department of Anesthesia and Intensive Care Medicine, Medical University Vienna; ClinicalTrials.gov Identifier: NCT01627613). As such, the TIP peptide represents a therapeutic candidate for the treatment of pulmonary edema, but its mechanism of action remains elusive. This study was designed to determine how the TIP peptide, mimicking the lectin-like domain of TNF, activates ENaC in the presence or absence of pneumolysin (PLY). We also investigated whether the lectin-like domain of TNF plays a physiological role in ALC during PLY-induced edema formation. Some of the results of these studies have been previously reported in the form of an abstract (37).
containing the TIP domain, located in the first exon, was subcloned into pBluescript II KS (Stratagene, La Jolla, CA), as such generating the pBluescript mTNF subconstruct. Desired point mutations were inserted making use of the QuikChange Site-Directed Mutagenesis Kit (Agilent Technologies, Santa Clara, CA) in combination with appropriate primers. Molecular characterization of positive embryonic stem (ES) cell clones required insertion of a Leu110Ser mutation in the third round, to generate an additional NheI restriction site. Next, the modified PstI/HindIII fragment was reintegrated into the sequence for native TNF. A neomycin resistance/thymidine kinase (neo/TK) selection cassette, flanked by loxP sites (fl), was inserted into the mutated TNF construct to select transformed mouse ES cells. The sequence of the selection cassette was isolated from the vector construct and was integrated at nucleotide position 9,552 into an intron region of the TNF gene. This integration of the triple-mutated gene into the mouse genome was performed at the Roche Centre for Medical Genomics in the laboratories of Dr. Horst Bluethmann (F. Hoffmann-La Roche AG, Basel, Switzerland). The targeting vector was first introduced by electroporation into a culture of C57BL/6-derived ES cells. Cells in which the mutated gene, including the selection cassette, was integrated into the genome upon homologous recombination were selected for resistance to the neomycin-like drug G418. Subsequently, cells were transfected with Cre recombinase, an enzyme recognizing loxP sites, which excises the intervening DNA. Cre/lox recombination caused removal of the selection cassette from the mutated gene. Cells were then injected into mouse blastocysts. Reimplantation of these blastocysts into the uterus of pseudo-pregnant dams resulted in the birth of 27 chimeric mice, 5 of which (4 female, 1 male) were intercrossed with C57BL/6 mice. Genotyping demonstrated that the first generation of offspring from the chimeric mice included heterozygous individuals for the mutated gene, implying that the ES cells had entered the germline of the chimeric animals.

Results

The Primary Target of the TIP Peptide Is ENaC

Because both Na\(^{+}\)-K\(^{+}\)-ATPase (29) and ENaC (27, 28, 38) can be activated by the TIP peptide, we investigated in more detail which of these Na\(^{+}\) transporters is the primary target in a homologous cell system and whether the peptide has a direct or indirect effect on their activity.

![Figure 1](image-url)

Figure 1. (A) Whole-cell, voltage-clamped patch-clamp current measurements (pA) of H441 cells treated or not with TIP peptide (50 μg/ml) in the cell bath in the presence or absence of amiloride (10 μM, n = 3/group). Inset: Effect of N-glycosidase F pretreatment (100 U, 5 min) of H441 cells on TIP peptide–mediated increase in Na\(^{+}\) uptake. (B–D) Single-channel patch-clamp measurements of 2F3 cells treated for 10 minutes with TIP peptide (50 μg/ml, n = 5) (B) or with TIP peptide pretreated with N,N'-diacetylchitobiose (100 μg/ml, n = 6) (C) or cellobiose (100 μg/ml, n = 6) (D).
As shown in Figure 1A, TIP peptide (50 μg/ml), when applied in the cell bath, significantly increased inward currents in H441 cells, a model of human Na⁺ absorptive airway epithelia (39), using a whole-cell, voltage-clamped, patch-clamp protocol with voltage steps ranging from -140 to +60 mV. The recorded current was completely inhibited by the ENaC inhibitor amiloride (10 μM).

Deglycosylation of H441 cells with 100 U of N-glycosidase F 5 minutes before adding TIP peptide blunted the peptide's effects on Na⁺ uptake (Figure 1A, inset), suggesting a crucial interaction of the peptide with glycosylated plasma membrane components, in correspondence with our previously obtained data in a heterologous cellular ENaC expression system (38).

To further test the effect of the TIP peptide on amiloride-sensitive ENaC activity, we performed single-channel analysis in membrane patches from Xenopus 2F3 cells (in cell-attached mode). As shown in Figure 1B, P₀ of ENaC increased from 0.18 ± 0.03 (before addition of TIP peptide) to 0.30 ± 0.04 (10 minutes after addition of TIP peptide into the pipette; P < 0.03; n = 5). Addition of the mutant TIP peptide, in which three residues (one Thr and two Glu) crucial for the Na⁺ uptake activation were exchanged against Ala (27), had no stimulatory effect (10 minutes after addition of mutant TIP: P₀ = 0.15 ± 0.04; n = 5; data not shown).

Preincubation of TIP peptide with a sugar specifically binding to the lectin-like domain of TNF (N,N'-diacetylchitobiose (100 μg/ml)) (25, 26) decreased P₀ from 0.22 ± 0.04 to 0.10 ± 0.03 (10 minutes after addition of TIP; n = 6) (Figure 1C), whereas the control sugar cellobiose (100 μg/ml) had no inhibitory effect (P₀ increased from 0.20 ± 0.03 [before addition] to 0.29 ± 0.04 [10 minutes after addition] of TIP; n = 6) (Figure 1D).

Pull-down experiments in which we used biotinylated human TNF (hTNF) revealed direct binding of human TNF (10 ng/ml) to recombinant ENaC-α, but not to the β and γ subunits (Figure 2A). A 2,000-fold molar excess of a PEGylated, soluble TNF receptor 1 complex over hTNF (20 μg/ml; Amgen, Thousand Oaks, CA), which does not interfere with the ALC-activating effect of TNF (21), only moderately blunted binding. By contrast, N,N'-diacetylchitobiose (100 μg/ml), but not cellobiose at the same concentration, completely inhibited the interaction (Figure 2A). In support of these results, there was strong binding activity of the TIP peptide, mimicking the lectin-like domain of TNF, to recombinant ENaC-α, with much weaker binding to the β and γ subunits (Figure 2B). As demonstrated in

Figure 2. (A) Representative pull-down experiment assessing binding of biotinylated human TNF (10 ng/ml) to the apical epithelial Na⁺ channel (ENaC)-α, -β, and -γ subunits in H441 cell lysates. (B) Measurement of binding of biotinylated TIP peptide to recombinant ENaC-α, -β, and -γ subunits (n = 5). (C) Upper panel: Representative pull-down experiment assessing binding of biotinylated TIP or mutant TIP peptide to endogenous or overexpressed ENaC-α in H441 cell lysates. Lower panel: Influence of preincubation of TIP peptide with either N,N'-diacetylchitobiose or cellobiose (both in 10-fold molar excess over TIP peptide) upon binding to ENaC-α. (D) Binding activity of biotinylated TIP or mutant TIP peptide to glutathione-S-transferase–coupled domains of ENaC-α. EL loop = extracellular loop.
involved in the Na
that oligosaccharide binding to TNF
Na study of molecular docking between TNF
cellobiose had no effect.

These results correspond to data from a study of molecular docking between TNF and Na
-diacetylchitobiose, indicating that oligosaccharide binding to TNF sterically hinders its association with residues involved in the Na⁺ uptake stimulating effect of the cytokine (40).

As shown in Table 1, the TIP peptide (50 µg/ml) and the mutant TIP peptide failed to increase the enzymatic activity or the Na⁺-binding affinity of the Na⁺-K⁺-ATPase at either pH 7.5 or pH 6.5. These data indicate that TIP peptide concentrations previously shown to activate Na⁺ uptake (27) do not directly modify Na⁺-K⁺-ATPase function.

**TIP Peptide Directly Binds to the Carboxy-Terminal Domain of ENaC-α**
To determine to which subdomain of ENaC-α the lectin-like domain of TNF binds, we used glutathione-S-transferase–tagged recombinant proteins of extracellular loop, amino-terminal, and carboxy-terminal domains of ENaC-α (14). Proteins were coupled to glutathione-S-transferase SpinTrap columns (GE Healthcare, Piscataway, NJ) and incubated with biotinylated or mutant TIP peptide. After elution of the complexes, biotinylated TIP peptide was detected only in the eluent of the carboxy-terminal domain–containing column (Figure 2D). Molecular docking studies carried out with the homology modeling approach and using the structure of the ENaC-related, acid-sensing ion channel 1 (ASIC-1) as a template (41) predicted a preferential binding of the TIP peptide, but not of the mutant TIP peptide, to the carboxy-terminal domain of ENaC-α, in correlation with our experimental data (Figure E1 in the online supplement). The potential interaction domain between the lectin-like domain of TNF and ENaC-α was previously proposed to be involved in gating (42).

The observation that the TIP peptide interacts with the carboxy-terminal intracellular domain of ENaC-α suggests it has to be internalized. Figures 3A and 3B demonstrate binding and uptake of rhodamine-labeled TIP peptide in H441 cells, reaching a maximal uptake level around 60 minutes, as quantified in Figure 3C. This uptake was completely blunted upon depletion of cholesterol in the H441 cells by 1 mM methyl-β-cyclodextrin (Figures 3A–3C). Cholesterol depletion has been shown not to affect ENaC expression, but rather to reduce its P₀ time and to induce its redistribution from low-density, cholesterol-rich membrane rafts to higher-density regions of the membrane (43, 44). As such, these data indicate that the TIP peptide has to be taken up in cholesterol-rich lipid rafts in the vicinity of ENaC.

**The Lectin-like Domain of TNF Blunts PLY-induced Down-regulation of ENaC Expression**
Toxins of bacterial and viral origin, such as LPS, PLY, and influenza A virus M2 protein, can significantly impair ENaC activity in a p38 MAP kinase– or PKC-dependent manner (33–36). We have recently shown that the pneumococcal toxin PLY impairs ENaC function (34). As shown in Figure 4A, PLY (60 ng/ml) reduces ENaC-α protein expression in H441 cells after 24-hour incubation, an effect blunted by coinoculation of the cells with TIP peptide (50 µg/ml). Interactions between P³ motifs in the carboxy-terminal domain of human ENaC subunits with WW domains of the E3 ubiquitin ligases Nedd-4 ½ and Nedd-4 ½ are facilitated by extracellular signal-regulated protein kinases 1 and 2 (ERK1/2)–mediated phosphorylation of T residues in the vicinity of the P³ motif, which initiates its degradation (11, 45). ERK1/2 is in turn activated by the phospholipase C (PLC)/PKC pathway (46). The protective effect of the peptide on ENaC-α expression likely occurs by means of inhibiting the PLC/PKC/ERK1/2 pathway. Indeed, the TIP peptide blunts PLY (125 ng/ml)–mediated activation of PLC (Figure 4F), PKC-α (Figures 4B and 4C) and ERK1/2 (Figures 4D and 4E). The PKC-α-specific inhibitor Ro32-0432 (10 nM) blunted the PLY-mediated suppression of ENaC-α protein expression (Figure 4G). This finding suggests an important role for this PKC isoform in PLY-mediated ENaC dysfunction. In the presence of PLY, the PKC-α inhibitor Ro32-0432 induces activation of PKC-δ (Figure 4H), an effect which does not occur with the TIP peptide (Figure 4I). A plausible explanation for the differential effects of a PKC-α inhibitor versus the TIP peptide on the status of PKC-δ in PLY-treated cells is that the peptide blunts PLC activation, which is upstream from both PKC-α and PKC-δ. The ENaC inhibitor amiloride (10 µM) partially abolished the protective activity of the TIP peptide on PLY-mediated PKC-α activation, which indicates cross-talk between ENaC and PKC-α, as previously proposed by others (47; see also Figures 4B and 4C).

**The Lectin-like Domain of TNF Preserves ENaC Open Probability in the Presence of PLY by Stabilizing the ENaC-MARCKS-PIP2 Complex**
Because the P₀ of ENaC is at least partially regulated by its complex formation with PIP2 and MARCKS (12–14), we assessed the influence of the TIP peptide on the ENaC–MARCKS association in the presence of PLY. As shown in Figures 5A and 5B, a 30-minute treatment of 2F3 cells with PLY (60 ng/ml) led to a significantly reduced association of ENaC-α with MARCKS, as compared to vehicle-treated cells. A 10-minute pretreatment with the
TIP peptide (50 μg/ml) significantly preserved the ENaC–MARCKS association in PLY-treated cells. Together with our findings that the TIP peptide preserves ENaC-α protein expression in the presence of PLY, these data provide a rationale for the capacity of the peptide to increase ENaC activity in the presence of the pneumococcal toxin.

TKI Mice Are More Susceptible to PLY-induced Impairment of ALC Than Wild-Type Mice

Up to 10% of pneumococcal pneumonia patients succumb days after their lungs are sterile following antibiotic treatment (48). A plausible explanation for this mortality is the massive release in the alveoli of the pneumococcal cholesterol-binding, pore-forming toxin PLY, an important mediator of permeability edema (48–50). In contrast to LPS, PLY does not induce a strong inflammatory response (51). We recently demonstrated that PLY impairs ENaC function in H441 cells (34).

To investigate whether the lectin-like domain of TNF can play a physiological role in ALC, we compared edema formation after intratracheal instillation of a low dose of PLY between wild-type (WT) and TKI C57BL6 mice, which produce a mutant TNF with a deficient Na⁺ uptake-activating capacity (23; see also Methods section). We selected a dose of PLY (1.5 μg/kg) that does not significantly impair alveolar epithelial and capillary barrier function, as shown in Figures 6A and 6C. Within 24 hours, this dose of PLY (1.5 μg/kg) caused a modest but significant increase in the generation of the proinflammatory cytokines IL-6 and TNF in the bronchoalveolar lavage fluid (Figure 6B). In the absence of barrier dysfunction (Figures 6A and 6C) and increased cell infiltration (data not shown), resident alveolar macrophages or AECs are the likely source of these mediators. The increase in TNF generation induced by PLY is comparable between WT and TKI mice, which allowed us to compare the actions of WT TNF, generated in the WT animals, and T104A-E106A-E109A mutant TNF in the TKI mice, on ALC, as assessed as lung wet-to-dry ratio. The lung wet-to-dry ratio was significantly higher in TKI mice than in WT mice 24 hours after PLY instillation (Figure 6D). In lung homogenates from TKI mice, we detected a more important reduction in protein expression of ENaC-α, but neither the β nor γ ENaC subunits, than in WT animals upon PLY treatment (Figures 6E and 6F).

Discussion

In previous studies, our research group and other investigators have demonstrated that the lectin-like domain of TNF represents a potent stimulator of ALC in several mammalian species and involves activation of ENaC via an unknown mechanism (21, 24, 27–31, 38). This study was designed to determine the mechanism by which the lectin-like domain of TNF can promote ENaC activity. We also investigated whether the lectin-like domain of TNF plays a physiological role in ALC during infection and inflammation. The results of this study demonstrate, for the first time to our knowledge, that ENaC activation by the TIP peptide involves multiple steps, starting with the interaction with glycosylated membrane components, possibly within the extracellular loops of ENaC (38) (Figure 1), followed by uptake within cholesterol-rich lipid rafts (Figure 3) and finally binding to the carboxy-terminal domain of the α subunit (Figure 2). A similar, multistep recognition and action
Figure 4. (A) Effect of pneumolysin (PLY) (60 ng/ml, 24 h) treatment on apical epithelial Na⁺ channel (ENaC) expression in H441 cells, assessed in a representative Western blot (WB). (B) Representative WB used to demonstrate protein kinase C-α (PKC-α) activation, measured as the ratio of phosphorylated (phospho)Ser657/total PKC-α expression in the plasma membrane in H441 cells treated for 1 hour with PLY (60 ng/ml). Cells were pretreated for 30 minutes with TIP peptide (50 μg/ml), either in combination with amiloride (10 μM) or not. (C) Quantification of the phospho/total PKC-α ratio in H441 cells treated with PLY in the presence or absence of TIP peptide (50 μg/ml) and amiloride (10 μM) (mean ± SD data from three independent experiments in triplicates). (D) Representative WB used to assess activation of extracellular signal-regulated protein kinases 1 and 2 (ERK1/2) in H441
Figure 5. (A) Representative immunoprecipitation Western blot showing that the association between apical epithelial Na<sup>+</sup> channel (ENaC) and myristoylated alanine-rich C kinase substrate (MARCKS) is attenuated after treating Xenopus 2F3 cells with pemoxilinol (PLY) (60 ng/ml) for 30 minutes, in comparison to vehicle-treated cells. The reduced association between ENaC and MARCKS was less pronounced in cells treated with TIP (50 µg/ml) 5 minutes before PLY treatment. WCL = whole-cell lysate. (B) Densitometry of immunoreactive bands corresponding to the ENaC-α subunit from three independent experiments. All values were normalized to the background signal.

Figure 4. (Continued), cells treated for 1.5 hours with PLY (125 ng/ml) upon pretreatment or not with TIP peptide (50 µg/ml, 30 min). (E) Quantification of the phospho/total ERK1/2 ratio in H441 cells treated with PLY in the presence or absence of TIP peptide (mean ± SD of three independent experiments in triplicates). (F) Effect of preincubation with the phosphatidylcholine-specific phospholipase C (PC-PLC) inhibitor D609 (30 µM) or with TIP peptide (50 µg/ml) upon PC-PLC activation in basal or PLY-treated H441 cells. (G) Ro32-0432 (10 nM) preserves ENaC-α expression in the presence of PLY (125 ng/ml) in H441 cells. (H) Influence of the PKC-α inhibitor Ro32-0432 (10 nM, 30 min) upon PKC-δ activation in H441 cells in the presence or absence of PLY (125 ng/ml). (I) Effect of TIP peptide pretreatment on PKC-δ and ERK1/2 activation in control and PLY-treated H441 cells.

Czikora, Alli, Bao, et al.; Novel Mechanism of Direct ENaC Activation

529
not induce significant barrier dysfunction in the lungs. Yet, this dose allowed us to specifically investigate the effect of TIP on PLY-induced impairment of ALC rather than on barrier function. Moreover, we have previously shown that the TIP peptide possesses barrier-protective effects in permeability edema induced by instillation of higher doses of PLY (30).

Our approach to increasing lung liquid clearance by means of directly inducing ENaC activity in a CAMP-independent manner can potentially provide an interesting alternative treatment option for pulmonary edema. Limitations of this approach could come from conditions in which there is excessive damage of the alveolar epithelium, such that insufficient ENaC is expressed on their apical surface to be activated by the TIP peptide. Moreover, excess protein deposition in the alveolar compartment during the recovery phase from alveolar edema can potentially occur because the salt and water fraction of the edema fluid are cleared much faster than protein (55).

Taken together, our data suggest that the lectin-like domain of TNF activates ENaC by means of a unique mechanism involving direct binding to the ENaC-\(\alpha\) subunit. This is, to our knowledge, the first example of a direct interaction between a cytokine and ENaC. The TIP peptide, mimicking the lectin-like domain of the cytokine, has been demonstrated to have promising protective effects in both preclinical and clinical settings of acute lung injury–associated edema, which should provide motivation for further evaluation of it as a potential novel therapeutic candidate for the treatment of pulmonary edema.

** AUTHOR disclo**

s are available with the text of this article at www.atsjournals.org.

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References


41. Stockand JD, Staruschenko A, Pochynyuk O, Booth RE, Silverthorn DU. Insight toward epithelial Na⁺ channel mechanism revealed by the acid-sensing ion channel 1 structure. IUBMB Life 2008;60:620–628.


