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Abstract

Minor protease inhibitor (PI) mutations often exist as polymorphisms in HIV-1 sequences from treatment-naïve patients. Previous studies showed that their presence impairs the antiretroviral treatment (ART) response. Evaluating these findings in a larger cohort is essential.

Reference


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Minor Protease Inhibitor Mutations at Baseline Do Not Increase the Risk for a Virological Failure in HIV-1 Subtype B Infected Patients

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Abstract

**Background:** Minor protease inhibitor (PI) mutations often exist as polymorphisms in HIV-1 sequences from treatment-naïve patients. Previous studies showed that their presence impairs the antiretroviral treatment (ART) response. Evaluating these findings in a larger cohort is essential.

**Methods:** To study the impact of minor PI mutations on time to viral suppression and time to virological failure, we included patients from the Swiss HIV Cohort Study infected with HIV-1 subtype B who started first-line ART with a PI and two nucleoside reverse transcriptase inhibitors. Cox regression models were performed to compare the outcomes among patients with 0 and ≥1 minor PI mutation. Models were adjusted for baseline HIV-1 RNA, CD4 cell count, sex, transmission category, age, ethnicity, year of ART start, the presence of nucleoside reverse transcriptase inhibitor mutations, and stratified for the administered PIs.

**Results:** We included 1199 patients of whom 944 (78.7%) received a boosted PI. Minor PI mutations associated with the administered PI were common: 41.7%, 16.1%, 4.7% and 1.9% had 1, 2, 3 or ≥4 mutations, respectively. The time to viral suppression was similar between patients with 0 (reference) and ≥1 minor PI mutation (multivariable hazard ratio (HR): 1.1 [95% confidence interval (CI): 1.0–1.3], P = .765). The time to virological failure was also similar (multivariable HR: 0.9 [95% CI: 0.5–1.6], P = .765). In addition, the impact of each single minor PI mutation was analyzed separately: none was significantly associated with the treatment outcome.

**Conclusions:** The presence of minor PI mutations at baseline has no effect on the therapy outcome in HIV infected individuals.


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Introduction

Minor protease inhibitor (PI) mutations are very common among treatment-naïve patients infected with HIV-1 but their impact on treatment outcome is poorly understood [1,2,3,4,5,6]. The prevalence of different minor PI mutations among treatment-naïve patients varies largely and is highly dependent on the HIV-1 subtype [7,8,9]. Some minor PI mutations occur as natural polymorphisms whereas others do not occur in the absence of PI therapy [10]. Minor PI mutations do not lead to high level resistance when occurring alone but they either improve the viral fitness or increase the drug resistance level in the presence of major PI mutations [11,12]. Minor PI mutations are therefore also called secondary or accessory mutations [13].

It was assumed that minor PI mutations among treatment-naïve patients might facilitate the emergence of major PI mutations and therefore lead to a worse therapeutic response to PIs. Other studies analyzing this issue were quite controversial. Perno et al. found evidence that the presence of minor PI mutations, particularly at position 10 and 36, lead to early treatment failure and to a higher number of acquired major PI mutations at the time of treatment failure [14,15]. Other studies found no evidence for an impaired treatment outcome [16,17,18,19]. All these studies are limited by a rather small sample size and mainly focus on response to unboosted PI therapy which is no longer recommended [20].

Therefore, we aimed studying the impact of minor PI mutations on virological outcome in first-line antiretroviral therapy (ART) using the dataset of the Swiss HIV Cohort Study (SHCS) [21].

Methods

Ethics Statement

The SHCS has been approved by the following ethical committees of all participating institutions: Kantonale Ethikkommission Bern; Ethikkommission beider Basel; comité d’éthique du département de médecine de Hôpitaux Universitaires de Genève; commission d’éthique de la recherche clinique, Lausanne; comitato etico cantonale, Bellinzona; Ethikkommission des Kanton St.Gallens; and Ethik-Kommission Zurich, all Switzerland. Written informed consent has been obtained from all participants.

Study Population

We used data from the SHCS, a nationwide, multicenter, clinic-based cohort with continuous enrolment and semi-annual study visits. Data up to 13 September 2011 were considered. The SHCS is very representative and includes about 66% of patients living with AIDS in Switzerland and 75% of all patients receiving antiretroviral therapy [21]. In addition, we used data from the SHCS drug resistance database that includes sequences from all authorized laboratories in Switzerland. Sequences are stored in SmartGene’s (Zug, Switzerland) Integrated Database Network System (IDNS version 3.6.5) [22].

Patient Selection and Study Design

We included HIV-1 subtype B infected individuals who started first-line ART between 1 January 1999 and 1 July 2010 with an unboosted PI or a boosted PI and two nucleoside reverse transcriptase inhibitors (NRTIs) and who had CD4 cell counts and HIV-1 plasma RNA levels measured before start of ART. A genotypic resistance test performed while ART-naïve was an additional inclusion criterion. Patients were excluded if they had viruses with ≥1 transmitted major PI mutation or if they had no HIV-1 RNA measured during first-line ART [23].

We studied the following endpoints: a) time to viral suppression, b) time to virological failure, and c) accumulation of major mutations at the time of virological failure. Time to viral suppression was defined as the time to the first viral load <50 copies/mL. Virological failure was defined as 2 consecutive values >500 copies/mL after at least 180 days of continuous treatment, 1 value >500 after 180 days followed by a treatment change or no viral suppression for more than 180 days. To fulfill the criteria of a virological failure, patients needed a minimum time of follow-up, therefore the analysis of time to virological failure was restricted to patients with ≥1 HIV-1 RNA measurement after 180 days of continuous treatment or to patients with ≥1 HIV-1 RNA measurement after previous viral suppression. The accumulation of major mutations at virological failure was studied in patients who experienced a virological failure on first-line ART and who had a genotypic resistance test performed between the virological failure and treatment change.


Statistical Analysis

We performed Fisher’s exact tests and Wilcoxon rank sum tests to compare categorical and continuous baseline characteristics, respectively. We plotted Kaplan-Meier curves and used log-rank tests to compare the virological outcome between patients with and without minor PI mutations. In addition, we performed univariable and multivariable Cox regression to analyze the time to viral suppression and the time to virological failure. Multivariable models were adjusted for the following potential confounders: sex, ethnicity, age, transmission category, baseline CD4 cell count, baseline HIV-1 RNA level, calendar year of ART start and the presence of NRTI mutations [23] and stratified for the PI used. Continuous variables were categorized if likelihood ratio tests showed significant departure from linearity. Follow-up was censored when first-line ART was changed or stopped. We checked the proportional hazard assumption with Schoenfeld residuals and by using graphical methods. No violation was found.

We also studied the impact of specific minor PI mutations on virological outcome. Here, only mutations with a prevalence ≥5% were considered. Despite this restriction, the number of events for some mutations was quite small, particularly the number of virological failures. Therefore, we used other methods that can deal better with rare events. It was shown that propensity scores are a good alternative to control for imbalances between groups when there are only small numbers of events per
In a 2-step procedure, we first calculated for each patient the propensity of being in the group with or without minor PI mutation. This was done by calculating propensity scores with multivariable logistic regression models adjusted for baseline HIV-1 RNA level, baseline CD4 cell count, ethnicity, sex, transmission category, calendar year of ART start, presence of NRTI mutations and the PI used. We validated if the propensity scores balanced the differences between groups adequately. Therefore, we performed logistic regression models adjusted for the propensity score to test if there were still imbalanced co-variables that were significantly associated with a group after adjustment. No poorly balanced co-variables were found. We did not use c statistics for model building of propensity score methods because it might be inadequate [25,26]. In a second step, we used the propensity scores for regression adjustment. The virological outcomes were analyzed with a Cox regression models adjusted for the log-transformed propensity score as the single co-variable. The log transformation is necessary for the adjustment because the variance of propensity scores needs to be similar between patients with and without minor PI mutations [27,28].

Statistical analyses were performed with Stata 11 (StataCorp, College Station, TX).

### Table 1. Baseline characteristics.

<table>
<thead>
<tr>
<th>Characteristics</th>
<th>≥1 minor PI mutation</th>
<th>No minor PI mutation</th>
<th>P*</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Sex</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Male</td>
<td>640 (82.9%)</td>
<td>342 (80.1%)</td>
<td>.227</td>
</tr>
<tr>
<td>Female</td>
<td>132 (17.1%)</td>
<td>85 (19.9%)</td>
<td></td>
</tr>
<tr>
<td><strong>Ethnicity</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>White</td>
<td>701 (90.8%)</td>
<td>398 (93.2%)</td>
<td>.149</td>
</tr>
<tr>
<td>Other</td>
<td>71 (9.2%)</td>
<td>29 (6.8%)</td>
<td></td>
</tr>
<tr>
<td><strong>Transmission category</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Men who have sex with men</td>
<td>426 (55.2%)</td>
<td>213 (49.9%)</td>
<td>.020</td>
</tr>
<tr>
<td>Heterosexual</td>
<td>187 (24.2%)</td>
<td>111 (26.0%)</td>
<td></td>
</tr>
<tr>
<td>Intravenous drug use</td>
<td>129 (16.7%)</td>
<td>95 (22.3%)</td>
<td></td>
</tr>
<tr>
<td>Other</td>
<td>30 (3.9%)</td>
<td>8 (1.9%)</td>
<td></td>
</tr>
<tr>
<td><strong>HIV-1 RNA</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>&lt;10,000 copies/mL</td>
<td>133 (17.2%)</td>
<td>70 (16.4%)</td>
<td>.263</td>
</tr>
<tr>
<td>10,000–99,999 copies/mL</td>
<td>278 (36.0%)</td>
<td>174 (40.8%)</td>
<td></td>
</tr>
<tr>
<td>&gt;100,000 copies/mL</td>
<td>361 (46.8%)</td>
<td>183 (42.9%)</td>
<td></td>
</tr>
<tr>
<td><strong>CD4 cell count</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>&lt;200 cells/μL</td>
<td>334 (43.3%)</td>
<td>156 (36.5%)</td>
<td>.024</td>
</tr>
<tr>
<td>200–300 cells/μL</td>
<td>253 (32.8%)</td>
<td>141 (33.0%)</td>
<td></td>
</tr>
<tr>
<td>&gt;350 cells/μL</td>
<td>185 (24.0%)</td>
<td>130 (30.4%)</td>
<td></td>
</tr>
<tr>
<td><strong>Median [IQR] CD4 cells/μL</strong></td>
<td>223 [125.5–339.5]</td>
<td>255 [141–379]</td>
<td>.010</td>
</tr>
<tr>
<td><strong>HIV-1 RNA</strong></td>
<td>137 (17.8%)</td>
<td>65 (15.2%)</td>
<td>.264</td>
</tr>
<tr>
<td><strong>Median [IQR] log_{10} HIV-1 RNA</strong></td>
<td>33 (4.3%)</td>
<td>12 (2.8%)</td>
<td>.201</td>
</tr>
<tr>
<td><strong>NRTI mutation</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Administered PIs</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>unboosted PIs</td>
<td>136 (17.6%)</td>
<td>119 (27.9%)</td>
<td>&lt;.001</td>
</tr>
<tr>
<td>boosted PIs</td>
<td>636 (82.4%)</td>
<td>308 (72.1%)</td>
<td></td>
</tr>
</tbody>
</table>

*P* Fishers exact p value for categorical variable and Wilcoxon rank sum for continuous variables.

Abbreviations: ART, antiretroviral therapy; CDC, Centers for Disease Control and Prevention; IQR, interquartile range; NRTI nucleoside reverse transcriptase inhibitor; PI, protease inhibitor.

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Results

Study Population and Baseline Characteristics

In the SHCS, 1265 subtype B-infected patients started first-line ART with a PI and 2 NRTIs and had a resistance test performed while ART-naïve. Patients were excluded from analysis if they had major PI mutations detected (n = 1), missing baseline HIV-1 RNA levels or CD4 cell counts (n = 14), or no HIV-1 RNA follow-up before the first ART change (n = 51). Finally, 1199 of 1265 patients (94.8%) were included to study the time to viral suppression. In table 1, we showed the baseline characteristics. Minor PI mutations were highly prevalent and present among 772 (64.4%) patients. Slightly more patients with a minor PI mutation were treated with a boosted PI, 82.4% compared to 72.1% without minor PI mutation (P < 0.001). The median CD4 cell count was higher among patients without minor PI mutation, 255 cells/μL compared to 223 cells/μL (P = 0.010). The most common NRTI combinations were lamivudine/zidovudine (43.3%), emtricitabine/tenofovir (33.2%) and lamivudine/abacavir (9.6%), no differences were observed between patients with and without minor PI mutations. 869 patients (72.5%) had the required minimum follow-up time to study the time to virological failure. Baseline characteristics of excluded patients did not markedly differ except that excluded patients started ART earlier (median: 2005 compared to 2006, P = 0.006).

Prevalence of Specific Minor PI Mutations

The prevalence of the most common minor PI mutations related to the respective administered PI therapies is shown in Figure 1. L63P was the most common minor PI mutation, it was present among 351 of 618 (56.8%) patients before treatment with lopinavir. Followed by the atazanavir related mutation I93L (41.2%, n = 114/277), the atazanavir/saquinavir related mutation I62V (n = 84/288, 29.2%) and the indinavir/nelfinavir/saquinavir related mutation V77I (n = 85/313, 27.2%). L10I and M36I were found to be associated with a worse treatment in previous studies [14,15]. In our study, they occurred in 9.7% (n = 118/1218) and 13.6% (n = 79/579) samples, respectively. The following mutations had a prevalence of <5%: L10F (0.2%)/R (0%)/V (1.9%), V11I (0%), K20I (0%)/M (0.4%)/R (2.3%)/T (0%)/V (0%), L24I (0.1%), V32I (0%), L33F (0.4%)/I (1.4%)/V (2.9%), E34Q (0%), E36L (1.1%)/V (0.4%), M46I (0.2%)/L (0%), I47V (0%), G48V (0%), I54A (0%)/L (0%)/M (0%)/S (0%)/T (0%)/V (0.2%), I64M (2.5%)/L (2.5%), A71I (0%)/L (0%), G73A (0%)/C (0%)/S (0%)/T (0%), T74P (0%), V82A (0.2%)/F (0%)/I (1.1%)/S (0%)/T (0.2%), I84V (0%), L90M (1.0%), and I93M (0%). Overall, 41.7%, 16.1%, 4.7% and 1.9% of patients had 1, 2, 3 and 4 minor PI mutations related to first-line ART.

Virological Outcome

The time to viral suppression and the time to virological failure were similar between patients with and without minor PI
mutations (Figure 2). Results of log-rank tests suggested no relevant differences. As shown in Table 2, univariable and multivariable hazard ratios (HR) were 1.1 (95% CI: 0.9–1.2) and 1.1 (95% CI: 1.0-1.3) when comparing the time to viral suppression between patients with and without minor PI mutations. A HR below 1 would indicate a longer time to viral suppression among patients carrying viruses with a minor PI mutation. Also the time to virological failure was not significantly different between patients with and without minor PI mutations, univariable and multivariable HRs were 1.0 (95% CI: 0.6–1.9) and 0.9 (95% CI: 0.5–1.9), respectively. The risk for a virological failure would be increased among patients detected with a minor PI mutation if the HR was above 1.

Additionally, we studied the effect of specific minor PI mutations on the virological outcome. No specific minor PI mutation was associated with a worse treatment outcome (Figure 3). We studied all minor PI mutations with a prevalence $5%. The 95% confidence interval of HRs always included 1, meaning that no significant differences were observed between patients with and without minor PI mutations.

We performed a sensitivity analyses and ran separate models for ART with unboosted PI and boosted PIs. For the time to viral suppression, multivariable HRs were 1.0 (95% CI: 0.6–1.9) and 1.2 (95% CI: 1.0–1.4) for therapies with unboosted and boosted PI, respectively. For the time to virological failure, multivariable HRs were 1.1 (95% CI: 0.6–2.0) and 0.6 (95% CI: 0.2–1.6) for unboosted and boosted PI therapies, respectively.

Accumulation of Major PI Mutations
The accumulation of major PI mutations was not higher among patients detected with a minor PI mutation while ART-naive. Of 63 patients who experienced a virological failure on cART, the accumulation of major PI mutations was not higher among patients detected with a minor PI mutation while ART-naive.
first-line ART, 43 (68.3%) had a resistance test performed. 7/19 (36.8%) patients without minor PI mutations and 9/24 (37.5%) accumulated a major mutation, respectively (P = 1.000).

Discussion

We found that the presence of minor PI mutations did not influence the virological outcome of first-line ART in HIV subtype B infected individuals. The prevalence of some minor PI mutations was found to be very high. Therefore, it is of great value to know that these mutations did not exhibit a negative impact on therapy outcome. In our study, neither the time to viral suppression, nor the time to virological failure differed between patients with and without minor PI mutations. Moreover, the risk for the emergence of a major PI mutation was not increased.

Today, first-line ART often includes non-nucleoside reverse transcriptase inhibitors, especially in resource-limited settings. However, PIs may increasingly be needed as good alternatives, especially in the presence of transmitted drug resistance mutations which seem to be seriously on the rise in resource-limited settings [29]. Our findings disproved concerns that the high prevalence of minor PI mutations limits the use of PIs but it is to mention that we only focused on subtype B infections.

To our knowledge, this is the largest study analyzing the impact of minor PI mutations on treatment outcome. We were able to include 1199 treated patients from the highly representative dataset of the SHCS. Despite the large number of patients included, the sample size was too small to perform an analysis for some specific minor PI mutations. Therefore, we had to restrict the analysis to the most prevalent minor PI mutations and used the propensity score method. The regression adjustment with propensity scores is a good option when the number of exposed patients is large and the number of events small. This method has the advantage that the Cox regression only had to be adjusted for one co-variable, the propensity score. If too many variables are included in a regression model relative to the number of events, estimates can be incorrect [30]. However, for some specific minor PI mutations, the confidence intervals of the HRs were quite large, especially for the models studying time to virological failure. This indicates that the accuracy of some estimates is limited. Unfortunately, we lacked statistical power to compare different combinations of minor PI mutations. Although we found that the time to viral suppression was shorter for patients with 3 minor PI mutations compared to patients without mutations, we think that the small difference we observed has no clinical relevance or even may have occurred by chance, considering that the lower bound of 95% CI of the HR was very close to 1, namely 1.1.

Our study supports findings from previous smaller studies that found no negative impact of minor PI mutations on therapy outcome [16,17,18]. However, it stands in contrast with the so far largest published studies by Perno et al including 248 and 93 individuals, respectively [14,15]. They found a higher risk of virological failure among patients with mutations at position 10 and 36 and a higher accumulation of major PI mutations. In contrast to our study, Perno et al. included different HIV-1 subtypes and they did not adjust their models for ethnicity. Ethnicity, however, is potentially an important confounder as it was found to be associated with treatment outcome in other studies [31,32]. Furthermore, Perno et al. mainly studied ART regimens containing unboosted PIs whereas our sample mainly contains regimens with boosted PIs. However, our sensitivity analysis that exclusively included ART with unboosted PIs containing 255 patients also lacked evidence for an impact of minor PI mutations on treatment outcome.

We convincingly demonstrated that minor PI mutations have no effect on virological outcome in PI-containing first-line ART, at least in patients infected by HIV-1 subtype B.

Acknowledgments


Author Contributions

Performed the experiments: JB SY CC TK. Analyzed the data: AUS VvW BL. Contributed reagents/materials/analysis tools: HFG AC MC LE PLV EB HF. Wrote the paper: AUS HFG.
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