Small risk of osteoarticular infections in children with asymptomatic oropharyngeal carriage of Kingella kingae

CERONI, Dimitri, et al.

Abstract

The aim of this study was to evaluate the absolute risk for children younger than 4 years of age with asymptomatic oropharyngeal carriage of Kingella kingae to sustain an osteoarticular infection. The rate of K. kingae carriage in the oropharyngeal mucosa was 9% among healthy children, and the risk for an asymptomatic carrier to develop an osteoarticular infection due to K. Kingae was estimated to be lower than 1%.

Reference


DOI: 10.1097/INF.0b013e31825d3419
PMID: 22572754
that in our case, the source of his infection was likely to be from his mother because the child had no contact with animals. The infant’s mother reported a history of a persistent cough before the infant’s illness, and B. bronchiseptica has the ability to inhibit leukocyte function and to adhere to respiratory epithelial cells, which could explain the persistence in the lower respiratory tract of both individuals. Our assumption on the direction of transmission is speculative because the mother was not screened for Bordetella at the time in which the child had his first positive culture.

REFERENCES

Accepted for publication May 01, 2012.
From the *Pediatric Orthopaedic Service, University of Geneva Hospitals; 1Division of Clinical Epidemiology, University of Geneva Hospitals; 2Centre for Clinical Research, University of Geneva Hospitals and Faculty of Medicine; 3Clinical Microbiology Laboratory, Service of Infectious Diseases, University of Geneva Hospitals; and 4Genomic Research Laboratory, Service of Infectious Diseases, University of Geneva Hospitals, Geneva, Switzerland.

K. kingae, as described earlier. Recruited children were either hospitalized for clean surgery, were attending our orthopedic outpatient clinic or visiting the emergency department for minor problems; children of hospital staff were also screened. Exclusion criteria were the presence of invasive diseases and the administration of antimicrobial drugs during the previous months.

METHODS
Study Design and Patients
In this prospective, consecutive study, we collected the data of all children aged 6–48 months admitted to the emergency department of the Geneva Children’s Hospital (111-bed tertiary pediatric hospital) with a suspicion of osteoarticular infection between January 1, 2008, and December 31, 2011. Eligible subjects underwent a clinical evaluation and blood analysis, and magnetic resonance imaging was performed on patients presenting clinical or laboratory findings suggestive for an osteoarticular infection and analyzed for signs of infection according to established criteria. Arthrocentesis or bone aspiration was performed under fluoroscopy guidance in the operating room for cases where imaging suggested osteoarticular infection.

In parallel, healthy children aged between 6 and 48 months were screened for asymptomatic oropharyngeal carriage of K. kingae during the same period. Oropharyngeal specimens were obtained by rubbing a cotton swab on the tonsils and subsequently analyzed by real-time polymerase chain reaction (PCR) for K. kingae as described earlier. Recruited children were either hospitalized for clean surgery, were attending our orthopedic outpatient clinic or visiting the emergency department for minor problems; children of hospital staff were also screened. Exclusion criteria were the presence of invasive diseases and the administration of antimicrobial drugs during the previous months.
Analysis of aspirates included a Gram stain (aerobic/anaerobic) culture and PCR assays. All biologic samples (oropharyngeal swabs, synovial fluid, bone biopsy specimen, peripheral blood) were analyzed with a new real-time PCR assay specific to *K. kingae*. Each PCR analysis was performed in duplicate. If the specific *K. kingae* PCR assay was negative, osteoarticular samples were submitted to broad-range PCR assay.

The diagnosis of osteoarticular infection caused by *K. kingae* was established when cultures and/or specific PCR assays were positive for this microorganism in blood, synovial fluid or bone aspirate samples. Osteoarticular infection caused by other microorganisms was defined by the presence of bacterial growth other than *K. kingae* on synovial fluid, blood or bone aspirate cultures and/or by a positive result in the broad-range PCR. The diagnosis of presumed osteoarticular infection was assigned when joint fluid/bone samples were not obtained for diagnostic bacterial culture or PCR assay or when cultures were negative, but magnetic resonance imaging demonstrated conclusive signs of infection. The diagnosis of osteoarticular infection was excluded when there were no positive bacterial findings or signs of osteoarticular infection on magnetic resonance imaging, or when improvement occurred without treatment.

The protocol was approved by the local institutional review board (09-029R, Mat-Ped 09-008R), and the study was conducted in compliance with Good Clinical Practice guidelines and the Declaration of Helsinki.

**Statistical Analysis**

The probability to sustain an osteoarticular infection due to *K. kingae* following a positive oropharyngeal PCR for *K. kingae* is equal to the positive predictive value (PPV) of the oropharyngeal PCR assay. The PPV can be derived from the sensitivity and specificity of the oropharyngeal PCR assay and the prevalence of *K. kingae* infection in the target population (PPV = p × Se/[p × Se + (1−p)×(1−Spe)]), where p is the prevalence in the target population, Se is the sensitivity of the oropharyngeal PCR assay and Spe is the specificity of the oropharyngeal PCR assay. The 95% confidence interval (CI) of the PPV is derived by applying the formula to the bounds of the 95% CI for sensitivity and specificity (box method).

**RESULTS**

Between January 1, 2008, and December 31, 2011, 64 consecutive patients met the eligible criteria for osteoarticular infection. Among these patients, 45 had *K. kingae* osteoarticular infection confirmed by culture or positive PCR from blood, bone or joint aspirate samples. Three had positive cultures for other microorganisms: *Haemophilus influenzae* (1 case); methicillin-sensitive *Staphylococcus aureus* (1 case); and *Streptococcus pneumoniae* (1 case). No organism was detected in the remaining 16. Among the latter patients, 6 presented radiologic and clinical findings of spondylodiscitis, but did not undergo disc-vertebral puncture. Ten cases showed imaging findings consistent with osteoarticular infection, but both cultures and PCR assays were negative, possibly due to difficult access to the infection site and/or low bacterial load.

Using the demographic census of the Geneva district, which reported a mean number of 16,950 children aged between 6 and 48 months per year during the study period, we calculated that the estimated proportion of young patients with an osteoarticular infection due to *K. kingae* in 1 year was less than 1% (p = 11.25/16,950 = 0.0664%).

Sensitivity of oropharyngeal PCR assay was assessed in a sample of 27 patients with osteoarticular infection caused by *K. kingae* confirmed by positive PCR assays specific for this microorganism on blood, synovial fluid or bone aspiration. The oropharyngeal PCR assay was positive in all cases, yielding a sensitivity of 100% (95% CI: 87.2–100). The specificity of oropharyngeal PCR was assessed by screening 431 asymptomatic patients. The PCR was negative in 396 subjects, yielding a specificity of 91.9% (95% CI: 88.9–94.3).

The probability to sustain an osteoarticular infection due to *K. kingae* following a positive oropharyngeal PCR assay is equal to the PPV of the PCR assay and was calculated by applying the formula described in the Methods section. The PPV among the 6- to 48-month-old population of Geneva and its surroundings was 0.81% (95% CI: 0.52–1.14%).

**DISCUSSION**

*K. kingae* is frequently isolated from the upper respiratory tracts of young children with a colonization rate ranging from 3.5% to 17.5%,11,12 Children usually first acquire *K. kingae* after the age of 6 months,6 the colonization rate then increases among infants aged between 12 and 24 months and finally declines among older children and adults.6,11,13 The present epidemiologic study confirmed a similar respiratory carriage rate in a Swiss pediatric population (9%), suggesting that asymptomatic colonization of the upper respiratory tract by *K. kingae* is probably a worldwide phenomenon. Similar to microorganisms such as *S. pneumoniae, H. influenzae* type b or *Neisseria meningitidis*,13,14 *K. kingae* is able to penetrate the bloodstream, disseminate and invade distant organs.6,14 Asymptomatic colonization of the upper respiratory tract is thus a strategy for *K. kingae* to survive and occasionally spread into the bloodstream under certain circumstances. A recent report described 3 patients with invasive *K. kingae* infections in whom the identical genotype isolate was recovered from the pharynx and the bloodstream. This observation supports the hypothesis that upper respiratory tract colonization represents the first step in the pathogenesis of invasive disease.4

The correlation between asymptomatic carriage and the subsequent incidence of osteoarticular infection due to *K. kingae* remains unclear, even if prophylactic antibiotic administration has been recently recommended in an attempt to prevent further disease spreading during outbreaks of *K. kingae* among day-care center attendees.3,5,15 Nevertheless, previous investigations on the epidemiologic features of asymptomatic respiratory carriage have not demonstrated any correlation between the pattern of carriage of *K. kingae* and the incidence of invasive disease.3 The probability for asymptomatic children carrying *K. kingae* in the oropharynx to develop an osteoarticular infection due to this pathogen is probably lower than 1%. Oropharyngeal carriage of *K. kingae* does not imply the subsequent development of invasive infection and suggests that other cofactors may play a role in the pathogenesis of invasive *K. kingae* infections.3

**ACKNOWLEDGMENTS**

The authors wish to thank the children and their families for their willing cooperation to participate in this study. The authors also acknowledge the staff of the pediatric orthopedics service of the Department of Child and Adolescent for their assistance.

**REFERENCES**


**POSTVARICELLA THROMBOSIS—REPORT OF TWO CASES AND LITERATURE REVIEW**

Fatma Rabah, MSc,* Nagwa El-Banna, MSc,* Mohamed Abdel-Baki, MD,† Ismail Beshlavi, MSc,‡ Divina Macaraig, DCH,* Depali Bhuyan, MRCPCH,* Mohamed Al-Hinai, DCH,* Nawal Al-Mashaikhi, CBR,* Shah Mohamed Wasiuddin, DCH,* Eileen Tomas, FRCPC,* and Anil Pathare, MD§

Abstract: Varicella (chickenpox) is a common disease of childhood, caused by varicella-zoster virus. Postviral thromboembolism is a rare complication of varicella-zoster virus in childhood. We describe 2 children who developed lower limb deep vein thrombosis shortly after varicella infection, along with a review of 130 previously reported cases.

Key Words: varicella, chickenpox, thrombosis, deep venous thrombosis, protein S deficiency, antiphospholipid antibody

Accepted for publication April 26, 2012.

From the *Department of Paediatric Haematology Oncology, Royal Hospital, Muscat, Oman; †Department of Pediatrics, Section of Hematology-Oncology, Baylor College of Medicine, Houston, TX; and the ‡Departments of Child Health and §Haematology, Sultan Qaboos University Hospital, Muscat, Oman.

The authors have no funding or conflicts of interest to disclose.

Address for correspondence: Anil V Pathare, MD, FCPS, FIMSA, Senior Consultant Haematologist, Sultan Qaboos University Hospital, PO Box 35, PC 123, Al Khod, Muscat, Oman. E-mail: apv16@hotmail.com.

Supplemental digital content is available for this article. Direct URL citations appear in the printed text and are provided in the HTML and PDF versions of this article on the journal’s Web site (www.pidj.com).

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DOI: 10.1097/INF.0b013e31825c7993

Varicella-zoster virus (VZV) causes primary, latent and recurrent infections. The primary infection manifests as varicella (chickenpox) and results in establishment of a lifelong latent infection of the sensory ganglion neurons. Reactivation of the latent infection causes herpes zoster. Although often a mild illness in childhood, chickenpox can cause substantial morbidity and mortality in otherwise healthy children.

There have been few case reports of thrombotic complications including transient antiphospholipid syndrome, protein C and protein S deficiencies associated with varicella. Purpura fulminans is the most common thrombotic complication, and thromboembolism reported in association with varicella has been shown to be due to both arterial and venous thrombi, generally after the skin rash.

We report here 2 cases of postvaricella thrombosis and review the literature to explore the wide spectrum of presentations and the different underlying pathophysiologic mechanisms, treatments and outcomes.

**CASE REPORT**

Case 1

A 10-year-old girl presented with a 2-day history of increasing pain and swelling of the left leg. Two weeks before the present symptoms, she had fever and a characteristic chickenpox rash all over the body, lasting for a few days with subsidence of fever following scab formation. She had no previous history of bleeding tendency or thrombosis. The patient was not immobilized or incapacitated during the acute viral illness. Her siblings also had chickenpox. There was no history suggestive of collagen vascular disorder. The family history was negative for bleeding disorders, thrombosis or stroke, and the parents were not consanguineous.

On examination, she had low-grade fever of 38°C with otherwise normal vital signs. Her left leg was swollen, purplish, tender and warm with pitting edema and normal peripheral pulses. A healed maculopapular rash was observed all over the body with centripetal distribution. The rest of the examination was unremarkable.

**FIGURE 1.** A and B, Doppler ultrasound demonstrating thrombus (A—arrows) in left common femoral vein (CFV), the left superficial femoral vein (SFV) and popliteal vein (POP) and its resolution (B).

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